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COMPARATIVE STUDIES OF SOME METABOLITES IN CARNY LEAF GALLS WITH PONGAMIA PINNATA LEAF GALLS

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Abstract

Accepted Date: There are hundreds of unique galls caused by insects and 10/10/2012 mites. They are formed on a variety of plants and in a **Publish Date:** broad range of sizes, shapes, colors and textures. Galls may be found on leaves, stems, twigs, branches, trunks 27/10/2012 and roots. Some galls are common and abundant and **Keywords** easily noticed. Others are rare or less conspicuous. This Galls paper reports the quantitative estimation of some metabolites in leaf galls of Carya. The parameters Carva assayed were Total sugars and Proteins. Total sugar content is less and protein content is more in Carny leaf **Total sugars** galls when compared to the Pongamiapinnata leaf galls. **Proteins**

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INTRODUCTION

Carya belongs to a family Juglandaceae, in Greek Carya means nut. Carya is commonly known as Hickory. The genus includes 17-19 species of deciduous trees with pinnately compound leaves and big nuts. Five or six species are native to China, Indochina and India, 11 or 12 are from the United States, two to four are from Canada and four are found in Mexico. It grows well in both wet and dry areas, but prefers well-drained soils. The Hickory tree is also used for fire wood and the Hickory flavor it adds to smoke meats. It is a great tree in areas with high winds because of the tap roots that go down deep in the earth making the trees very wind resistant. Man and animals enjoy the nut from the Hickory tree. Carya sect.Sinocarya is called Asian hickories. Galls are seen on leaves and fruits. Galls may also provide the insect with physical protection from predators^{1, 2}. In Carya galls are seen on leaf and leaf stem. The Carya leaf stem gall phylloxera (phylloxeracaryaecaulis) also uses as a food source. Egg hatch in early spring and the galls quickly from around the developing insects. Phylloxera galls may damage weakened or stressed hickories,

but is generally harmless. Deformed leaves and twigs can rain down from the tree in the spring as squirres break off infected tissue and eat the galls, possibly for the protein content or because the galls are fleshy and tasty to the squirrels. Pongamia pinnata, locally known as Karanja, is a mangrove plant belonging to the family Fabaceae. It is a medium size glabrous tree attaining a height of around 18 meter and its habitat is in the littoral regions of Southeast Asia, Australia and Fiji.^{3, 4} In Ayurveda and Unani medicine, used as antiinflammatory, anti-plasmodia, antinoneceptive, anti- hyperglycemic, antilipodo oxidative, antidiarrheal, antiulcer, anti-hyper ammonic and anti-oxidant⁵. Traditionally its bark is used in pile; leaves are effective as medicated both and Rheumatic pains, and the seeds are used in hypertension, bronchitis, whooping cough, skin diseases and Rheumatic arthritis.⁶⁻⁸. Roots are used for cleaning gums, teeth and ulcers, also effective in gonorrhea^{9, 10}. Insect gall formation is uncommon and plants¹¹. specific to only certain Pongamiapinnataand Carya are such plants.

Research Article Srilakshmi P, IJPRBS, 2012; Volume 1(5): 331-337

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MATERIALS & METHODS

Young galled leaves of equal size were collected from the local areas of Hyderabad, Andhra Pradesh, India during August 2012. Biochemical studies from gall affected leaves were conducted. These studies included estimation of total soluble sugar and Protein by Phenol Sulphuric acid reagent method¹² and by Lowry method¹³ respectively.

Estimation of total soluble sugar

The amount of total soluble sugars was estimated by Phenol sulphuric acid reagent method (Dubois et al., 1951).

500mg each of fresh normal and fungal infected gall was homogenized with 10ml of 80% ethanol. Then each sample was centrifuged at 2000rpm for 15-20min. The supernatant were collected separately, to 1ml of alcoholic extract, 1ml of 5% phenol solution was added and mixed. Then 5ml of 96% sulphuric acid was added. Each tube was gently agitated during the addition of the acid and then allowed to stand in a water bath at 25-30C for 20minutes. The OD of the characteristic yellow orange color thus developed was measured at 490nm in a spectrophotometer. Simultaneously a standard curve was prepared by using known concentration of glucose. The amount of sugar was expressed as mg/g fresh weight of tissue.

Estimation of Protein

Quantitative estimation of protein was carried out by the method of Lowry *et al* (1951). 1 gm plant material was

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Research Article Srilakshmi P, IJPRBS, 2012; Volume 1(5): 331-337

homogenized with 10ml of 80% ethanol. The extract was centrifuged at 5000rpm for 5 min and the supernatant was discarded. 5% 10ml TCA (Trichloroacetic acid) or per chloric acid (PCA) was added to the residue and incubated at 80°C for 20 minutes. The pellet was centrifuged and the supernatant was discarded. Residue was washed with 10ml distilled water and again centrifuged. The supernatant was discarded. 2% 10ml sodium carbonate in 0.1N NaOH was added to the residue and incubated for an hour at 30°C.Again centrifuged and residue was discarded. To 1ml of sample extract 5ml of alkaline copper reagent was added and then allowed to stand for 10minutes. Each tube was gently agitated during the addition of 0.5ml of Phenol reagent and incubated at room temperature in the dark for 30minutes till blue color was developed. The OD of the characteristic blue was measured at 660nm in а spectrophotometer after setting for 100% transmission against the blank, standard curve was prepared by using known concentrated of protein.

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There was a substantial reduction in total soluble sugars in Carny leaf galls. The levels of Protein in gall tissue showed increase compared to Pongamiapinnata. These results are shown in graphs (A and B).Sugars and Proteins are very important biochemical nutrients for the plants. If abundantly existing, these are detected naturally by various other micro-organisms like bacteria, fungi, insects etc. So once infected, inside the host tissue these organisms utilize the excess nutrients present in the plant for their growth and sustenance¹⁴.

RESULTS AND DISCUSSION

CONCLUSION

Therefore, from the present investigation we can conclude galls cause a change in

Quantities of metabolites. So these studies may be useful for the future investigations.



* G –Gall * HL – Healthy Leaf * GL – Gall Removed Leaf - TABLE - A



* G –Gall * HL – Healthy Leaf * GL – Gall Removed Leaf - TABLE - B

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REFERENCES

1. Weis AE and A Kapelinski: Variable selection on Eurosta's gall size. II. A path analysis of the ecological factors behind selection. Evolution. 1994; 48: 734 – 745.

2. Graham N Stone and Karsten Schonrogge: The adaptive significance of insect gall morphology. TRENDS in Ecology and Evolution. 2003; 18(10): 512-522.

3. Chopra RN, Nayar SL and Chopra IC: Glossary I f Indian Medicinal Plants (Including the supplement). Council of Scientific and Industrial Research, CSIR Publications, New Delhi. C.S.I.R (Council of Scientific and Industrial Research). The Wealth India 11 vols. New Delhi. 1998.

4. Simin K, Ali Z, khaliq-Uz-Zaman SM and Ahmad VU: Structure and biological activity of a new rotenoid from PongamiaPinnata. Nat Prod Lett. 2002; 16: 351-357.

5. Punitha R and Manoharan S: Antihyperglycemic and antilipidperoxidative effects of Pongamiapinnata (Linn) Pierre flowers in alloxan induced diabetic rats. J. Ethnopharmacol. 2009; 105: 39-46. 6. Ballal M: Screening of medicinal plants used in rural Indian folk medicine for treatment of diarrhea. 2005.

 Tanaka T, linuma M, Yuki K, Fuji Y and Mizuno M: Flavonoids in root bark of Pongamiapinnata. Phyto chemistry 1992; 31: 993-998.

8. Carcache Blanco EJ, Kang YH and Park EJ: Constituents of the stem bark of Pongamiapinnat with the potential to induce quinine reductase. J. Nat.Prods. 2003; 66: 1197-1202.

9. Rastogi RP and Malhotra BN: Compendium of Medicinal Plants, Central drug Research Institute Lucknow and National Institute of Science Communication, New Delhi, 2001; 522-523.

10. Chauhan D and Chauhan JS: Flavonoid glycosides from Pongamiapinnata.Pharma Biol.2002; 40:171-174.

11. Dreger-JauffretF.and Short house JD: Diversity of galls- Inducing insects and their galls. In Biology of insect-induced galls, J.D

Available Online At www.ijprbs.com

Research Article Srilakshmi P, IJPRBS, 2012; Volume 1(5): 331-337	ISSN: 2277-8713 IJPRBS
Short house and Forphfritsch (Ed), Oxford	with the Folin-phenol reagent. J. Biol. Chem.
University press, New York. 1992: 8-33.	193 263-275.
12. Dubois M. K., Gilles J.K., Robers P.A. andSmith F. (1951): Calorimetric determinationof sugar and related substance. Anal. chem.26, 51-356.	14. Sheen S J and Anderson R A 1974 Comparison of Polyphenols and related enzym3w in the capsule and nodal tumor of Nicotiana plants; Can. J. Bot. 52 1379.

13. Lowery OH, Rose brough NJ, Farr AL and Randall RJ (1951). Protein measurement